

## The morphophysiological dormancy in *Amborella trichopoda* seeds is a pleisiomorphic trait in angiosperms

Bruno Fogliani<sup>1,2,\*</sup>, Gildas Gâteblé<sup>1</sup>, Matthieu Villegente<sup>2</sup>, Isabelle Fabre<sup>1</sup>, Nicolas Klein<sup>1,2</sup>, Nicolas Anger<sup>1</sup>, Carol C. Baskin<sup>3,4</sup> and Charlie P. Scutt<sup>5</sup>

<sup>1</sup>Institut Agronomique néo-Calédonien (IAC), BP 73 Port Laguerre, 98890 Païta, New Caledonia, <sup>2</sup>Laboratoire Insulaire du Vivant et de l'Environnement (LIVE)–EA 4243, University of New Caledonia (UNC), BP R4, 98851 Noumea, New Caledonia, <sup>3</sup>Department of Biology, University of Kentucky, Lexington, KY 40506, USA, <sup>4</sup>Department of Plant and Soil Sciences, University of Kentucky, Lexington, KY 40546, USA and <sup>5</sup>Reproduction et Développement des Plantes (RDP; UMR5667, CNRS-INRA-Université de Lyon), Ecole Normale Supérieure de Lyon, 46 allée d'Italie, 69364 Lyon Cedex 07, France  
\*For correspondence. E-mail fogliani@iac.nc

Received: 14 July 2016 Returned for revision: 5 September 2016 Editorial decision: 5 October 2016

- **Background and Aims** Recent parsimony-based reconstructions suggest that seeds of early angiosperms had either morphophysiological or physiological dormancy, with the former considered as more probable. The aim of this study was to determine the class of seed dormancy present in *Amborella trichopoda*, the sole living representative of the most basal angiosperm lineage Amborellales, with a view to resolving fully the class of dormancy present at the base of the angiosperm clade.
- **Methods** Drupes of *A. trichopoda* without fleshy parts were germinated and dissected to observe their structure and embryo growth. Pre-treatments including acid scarification, gibberellin treatment and seed excision were tested to determine their influence on dormancy breakage and germination. Character-state mapping by maximum parsimony, incorporating data from the present work and published sources, was then used to determine the likely class of dormancy present in early angiosperms.
- **Key Results** Germination in *A. trichopoda* requires a warm stratification period of at least approx. 90 d, which is followed by endosperm swelling, causing the water-permeable pericarp–mesocarp envelope to split open. The embryo then grows rapidly within the seed, to radicle emergence some 17 d later and cotyledon emergence after an additional 24 d. Gibberellin treatment, acid scarification and excision of seeds from the surrounding drupe tissues all promoted germination by shortening the initial phase of dormancy, prior to embryo growth.
- **Conclusions** Seeds of *A. trichopoda* have non-deep simple morphophysiological dormancy, in which mechanical resistance of the pericarp–mesocarp envelope plays a key role in the initial physiological phase. Maximum parsimony analyses, including data obtained in the present work, indicate that morphophysiological dormancy is likely to be a pleisiomorphic trait in flowering plants. The significance of this conclusion for studies of early angiosperm evolution is discussed.

**Key words:** Amborellaceae, *Amborella trichopoda*, evolution of seed dormancy, ANA-grade angiosperm, seed germination, morphophysiological dormancy, rudimentary embryo.

### INTRODUCTION

Seed dormancy can be defined as the retardation of radicle emergence (germination) under otherwise favourable conditions for germination (Bewley *et al.*, 2015). There are five major classes of seed dormancy: morphological, physiological, morphophysiological, physical and combinational, within each of which further sub-classes may be recognized (Baskin and Baskin, 2004). Among the five major classes of dormancy, physiological dormancy (PD) is characterized by the presence in the mature seed of a fully developed embryo, which takes >30 d following imbibition before the radicle emerges. In contrast, morphological dormancy (MD) is characterized by the presence in the mature seed of an incompletely developed embryo that, under appropriate conditions, begins to grow immediately following imbibition, but takes <30 d to complete development before the radicle emerges. In morphophysiological dormancy (MPD), a combination of PD and MD is present, delaying germination for >30 d.

As dormancy is a key trait for survival and fitness (Linkies *et al.*, 2010), the reconstruction of the evolution of dormancy mechanisms is of particular interest. In the angiosperms, such reconstruction work is now possible as studies performed over the last approx. 15 years (summarized at <http://www.mobot.org/MOBOT/research/APweb/>; Byng *et al.*, 2016) have provided a solid phylogenetic framework. According to the majority of phylogenetic studies, the orders Amborellales, Nymphaeales and Austrobaileyales, collectively termed the ANA grade, diverged sequentially from a remaining lineage from which all other living angiosperms, the euangiosperms, are descended. A slightly different evolutionary hypothesis, which has emerged from a few analyses (Qiu *et al.*, 2001, 2010; Xi *et al.*, 2014), places Amborellales and Nymphaeales together in a first-diverging angiosperm clade, with Austrobaileyales as a second-diverging lineage, sister to euangiosperms. The euangiosperms are composed of the major groups of eudicots and monocots, in addition to the moderately

sized magnoliid clade and the two smaller taxa of Chloranthaceae and *Ceratophyllum*.

As a result of the largely successful resolution of angiosperm phylogeny, analyses of the basally diverging ANA-grade lineages have been particularly informative for the reconstruction of character states at the base of the flowering plants. The likely earliest diverging ANA-grade order, Amborellales, contains *Amborella trichopoda* Baill. as its only living representative. This species is a dioecious shrub, endemic to the understory of humid tropical forests on the southern Pacific island of New Caledonia. The mature *A. trichopoda* embryo is short and broad (Bailey and Swamy, 1948; Tobe *et al.*, 2000), suggesting the possible presence of MD. Fourcade *et al.* (2015) refer to unpublished studies of germination and dormancy in this species, though no detailed work has previously been undertaken.

The likely second-earliest diverging ANA-grade order, Nymphaeales, contains three aquatic families, Nymphaeaceae, Cabombaceae and Hydatellaceae, of which the latter is indicated by molecular phylogeny to be basally diverging (Saarela *et al.*, 2007). Physiological dormancy has been reported in the Nymphaeaceae genera *Nymphaea* (six species studied) and *Nuphar* (one species studied) and in the Cabombaceae genus *Cabomba* (one species studied), as summarized by Willis *et al.* (2014). In contrast, seeds of *Trithuria* (three species studied), the only genus of Hydatellaceae, have MPD (Tuckett *et al.*, 2010). Interestingly, MPD in *Trithuria* contains an extreme morphological component as embryos in mature *Trithuria* seeds show very limited differentiation and consist of only about 18–23 cells, depending on the species (Tuckett *et al.*, 2010; Friedman *et al.*, 2012).

The third-diverging ANA-grade order, Austrobaileyales, contains three families of woody species, Austrobaileyaceae, Trimeniaceae and Schisandraceae. The Schisandraceae genera *Schisandra* (three species studied) and *Illicium* (five species studied) both have MPD (summarized by Willis *et al.*, 2014), while dormancy in the remaining genera of Austrobaileyales has yet to be assessed.

A recent large-scale study of seed dormancy types (Willis *et al.*, 2014) has collated data on 13 939 angiosperm and 297 gymnosperm species, and concluded the likely ancestral state of dormancy in the seed plants to be MPD. According to this study, the most recent common ancestor (MRCA) of the angiosperms analysed (which included some Nymphaeales and Austrobaileyales, but not Amborellales) had either MPD or PD. Similarly, the MRCA of euangiosperms (i.e. all living angiosperms, excluding the ANA grade) was not completely resolved between MPD and PD, though MPD was considered more likely.

In the present work, we focus on seed dormancy in the key ANA-grade species *A. trichopoda*, likely sister to all other living flowering plants. We perform detailed quantitative and microscopic observations on germinating seeds, coupled with studies of the water permeability of seed-covering layers and the effects on germination of the exogenous application of the plant hormone GA<sub>3</sub>. We conclude that the precise type of dormancy in *A. trichopoda* seeds corresponds to the non-deep simple level of MPD. We incorporate these data, together with recently published data on Hydatellaceae, into character-state reconstructions in the angiosperms and conclude from this analysis that the likely pleisiomorphic state of the living flowering plants was MPD.

## MATERIALS AND METHODS

### Fruit collection

Mature fruits of *Amborella trichopoda* were harvested during the season of natural dispersal from plants growing at an altitude of around 600 m on the Plateau de Dogny in the central part of Grande Terre (S21°37'21.6", E165°52'06.3"), New Caledonia. One batch, collected on 8 July 2008 and sown 7 d later, was used for dissection and observations on embryo development, while a second batch was harvested on 23 December 2008 and used for germination experiments, starting on 15 January 2009. Empty drupes were removed under vacuum, and fleshy parts of the remaining drupes were removed by gently rubbing fruits on a sieve under running water prior to further studies. Drupes without fleshy parts (DWFP) were stored prior to use in plastic zip-lock bags at room temperature (25 ± 2 °C).

### Physical characteristics of mature DWFP and seeds

To assess *A. trichopoda* for the possible presence of dormancy, we first studied the physical characteristics of its mature DWFP and seed *sensu stricto*. Immediately after harvesting four replicates of 100 DWFP and of 100 seeds *sensu stricto* extracted from the DWFP were weighed. Also, three replicates of 20 DWFP and 20 seeds *sensu stricto* were measured, and average dimensions were determined. The length and width of entire DWFP, as well as those that had been cut transversally or longitudinally, were measured using a dissecting microscope equipped with a micrometer. DWFP were mounted on scanning electron microscope specimen holders using double-sided carbon adhesive tape and carbon-metalized (10–15 nm), as described by Rabier *et al.* (2008). Observations and size measurements were made using a Philips XL 30 environmental scanning electron microscope. Moisture content of four replicates of 25 DWFP was determined gravimetrically (103 ± 2 °C for 17 ± 1 h), by the protocol of MacKay *et al.* (2002), observing standard procedures (ISTA, 1999).

### Embryo growth and germination

As the physical characteristics of *A. trichopoda* seeds suggested the presence of dormancy, we conducted preliminary germination assays to evaluate directly the presence of dormancy and gain insights into the overall class of dormancy present. The embryo length:seed length (E:S) ratio was determined for 50 freshly matured DWFP collected in July and cut longitudinally. Fresh DWFP were placed in four sowing trays (25 DWFP per tray) on a 1:1 perlite:peat substrate, covered with a 4 mm layer of vermiculite and incubated at 25 ± 4 °C (corresponding to the average diurnal temperature in *A. trichopoda*'s natural habitat) on a greenhouse sowing table. Intact and longitudinally cut DWFP were photographed after 93 d, and the E:S ratio was determined from 20 DWFP that were beginning to break open.

### DWFP permeability

To elucidate the basis of dormancy in *A. trichopoda* seeds, we tested the water permeability of the mature DWFP. The seed *sensu stricto* and pit (pericarp/sclerified mesocarp) from

three replicates of 20 DWFP were weighed before and after soaking for 72 h in distilled water and means were compared using the Student *t*-test. Seeds were imbibed in 1 % (w/v) aqueous methylene blue dye and dissected to determine the path of water entry, as described by Orozco-Segovia *et al.* (2007).

#### Scarification treatment and removal of endocarp and hard layer of mesocarp

To investigate the mechanical role of the DWFP in *A. trichopoda* seed dormancy, we compared the time course of germination in intact DWFP, DWFP scarified by acid treatment, and seed *sensu stricto* that had been excised from all surrounding tissues. A total of 100 DWFP were scarified for 90 min in 10 mL of concentrated sulphuric acid, rinsed with abundant tap water and soaked in three consecutive distilled water baths until all soft tissue remnants had been removed. Scarified and non-treated (control) DWFP were sown and incubated as described above, but under conditions of 95 % relative humidity generated using a fog system. DWFP were monitored for hypocotyl emergence twice weekly for 1 year.

After removal of fleshy tissues, DWFP were surfaced sterilized with 2 % chlorine bleach for 5 min, then rinsed three times over a 5 min period in sterile distilled water. Extraction of the seed *sensu stricto* from the endocarp and the hard inner part of the mesocarp (pit) was performed using a thin scalpel blade and tweezers, being careful not to damage the seed coat or seed. After isolation, seeds *sensu stricto* were surface-sterilized under a laminar hood using 2 % (w/v) bleach for 5 min and rinsed three times in sterile distilled water. Sterile seeds were sown in Petri dishes on two layers of germination paper moistened with sterile distilled water. A total of 100 seeds were sown (25 per dish, four replicates) and incubated at 25 °C under continuous light. The criterion used for germination was radicle protrusion to a length of 1 mm.

#### Effects of gibberellic acid on germination

To investigate physiological dormancy in *A. trichopoda* seed, we used a discriminatory test which relies on the effect of gibberellin treatment; moderate concentrations of exogenous gibberellic acid (GA<sub>3</sub>) are known to promote germination in seeds showing non-deep PD, but not deep PD (Baskin and Baskin, 2004). Non-scarified DWFP were placed in Petri dishes on germination paper moistened with 0, 50, 100, 500 or 1000 ppm GA<sub>3</sub>. A total of 100 DWFP were sown for each treatment (25 DWFP per dish, four replicates) and incubated at 25 °C under continuous light, and examined weekly for radicle emergence. Time of germination was recorded at radicle protrusion, and means were compared using the Student *t*-test.

#### Test of epicotyl MPD

Four types of non-deep simple epicotyl MPD and four of deep simple epicotyl MPD have been recognized (Baskin and Baskin, 2014). To determine if seeds of *A. trichopoda* have epicotyl MPD, we measured the time necessary for cotyledon emergence, following radicle emergence. Ninety-eight DWFP were incubated for 24 h in water and 100 DWFP in 50 ppm

GA<sub>3</sub> (Sigma-Aldrich), before being sown on trays at high humidity, as described above, and observations were made on radicle and cotyledon emergence every 2 or 3 d.

To determine whether GA<sub>3</sub> would promote cotyledon emergence (and thus whether a type of non-deep simple epicotyl MPD was present), GA<sub>3</sub> (50 ppm, Sigma-Aldrich) dissolved in lanolin (anhydrous wool fat; Probiotec, Australia) was applied with a paintbrush to 5 mm long rootlets, sparing the tip, of the 38 seeds which had germinated from the group of 98 seeds incubated in water (above). This stage was reached 1–2 d before secondary rootlet formation, and approx. 11 d after radicle emergence. The normality of the data distribution within each treatment group was assessed using the Shapiro–Wilk normality test ( $\alpha = 0.05$ ). Statistical comparisons of the time between radicle and cotyledon emergence was made using the Welch test ( $\alpha = 0.05$ ) for normal distributions and the Mann–Whitney Wilcoxon test ( $\alpha = 0.05$ ) for non-normal distributions.

#### Character reconstruction by parsimony

Cladograms representing two alternative topologies for the base of angiosperm phylogeny were manually created in Newick format, incorporating information from the Angiosperm Phylogeny Group III Website (Byng *et al.*, 2016) and from Xi *et al.* (2014), respectively, with additional data from Saarela *et al.* (2007) and Löhne *et al.* (2007). The internal phylogeny of Nymphaeaceae was simplified by representing *Nymphaea*, *Victoria*, *Euryale* and *Ondinea* as a polytomy as, according to the phylogeny of Löhne *et al.* (2007), *Nymphaea* is polyphyletic, one section being sister to *Victoria* + *Euryale*, while another includes *Ondinea*. Character states given in Tuckett *et al.* (2010), Willis *et al.* (2014) and obtained in the present work were mapped onto both cladograms in Mesquite (Maddison and Maddison, 2015) using parsimony reconstruction.

## RESULTS

#### Physical characteristics of mature DWFP and seeds

Physical characteristics of *Amborella trichopoda* seeds suggest the presence of dormancy. The mass of the *A. trichopoda* seed *sensu stricto* represented approx. 27 % of the mass of the mature DWFP, while seed length and width were 70 and 45 %, respectively, those of the mature DWFP (Table 1). The seed possessed a thin seed coat (Fig. 1B, E), surrounded by a hard, thick pit (Fig. 1A–D) composed of an endocarp and a sclerified layer from the mesocarp (Fig. 1D), previously described by Bobrov *et al.* (2005). Interestingly, X-ray microscopic analysis revealed an amorphous silicate-crystal layer located between these two parts of the pericarp (Fig. 1E). In freshly matured DWFP of *A. trichopoda*, the embryo, which was surrounded by a copious endosperm (Fig. 1B), was found to be arrested at the heart-shaped stage (Fig. 1F). At this stage, the E:S ratio was  $0.08 \pm 0.01$ .

The potential resistance to embryo growth of the pericarp–mesocarp envelope, including the newly discovered amorphous silicate-crystal layer, is compatible with the possible presence of PD mechanisms in *A. trichopoda*. In addition, the early arrest of embryo development and the low E:S ratio in the freshly matured seed are both compatible with the possible presence of

TABLE 1. Characteristics of *Amborella trichopoda* drupes without fleshy parts (DWFP) and of seeds *sensu stricto*

	Average weight (g $\pm$ s.d.)	Average length (mm $\pm$ s.d.)	Average width (mm $\pm$ s.d.)	Water content (% $\pm$ s.d.)	E:S ratio
For 100 DWFP	2.15 $\pm$ 0.08	5.98 $\pm$ 0.24	3.49 $\pm$ 0.11	8.70 $\pm$ 0.06	0.08 $\pm$ 0.01
For 100 seed <i>sensu stricto</i>	0.58 $\pm$ 0.06	4.19 $\pm$ 0.99	1.57 $\pm$ 0.37	8.99 $\pm$ 0.65	

E.S ratio, embryo length:seed length ratio.

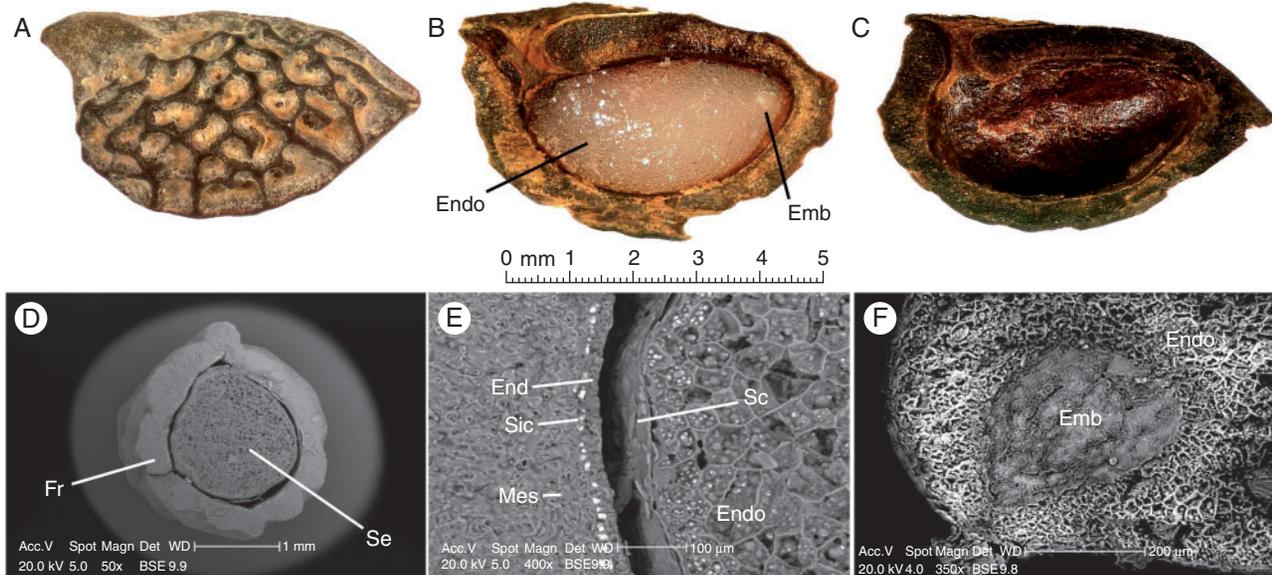


FIG. 1. *Amborella trichopoda* drupe without fleshy part (DWFP) structure. (A–C) Binocular microscopy, (D–F) scanning electron microscopy. (A) Whole DWFP. (B) Longitudinally cut DWFP. (C) An opened drupe showing the pit and seed *sensu stricto*. (D) Transverse section. (E) Magnified transverse section showing the meso-endocarp of the fruit separated by a silicate-crystal layer. (F) Longitudinal section of a seed *sensu stricto* showing the heart-shaped underdeveloped embryo (Emb, embryo; End, endocarp; Endo, endosperm; Fr, fruit parts; Mes, mesocarp; Sc, seed coat; Sic, silicate-crystal layer).

MD. We thus conducted preliminary germination assays to evaluate directly the presence of dormancy and gain insights into the overall type of any such mechanisms present.

#### Embryo growth and germination

A preliminary time course of embryo growth and germination indicated that *A. trichopoda* seeds have a level of simple MPD. The pit (endocarp and hard part of the mesocarp) was observed to split open after a minimum of approx. 90 d incubation at 25 °C. At this stage, the embryo was small and heart shaped (rudimentary), as in the freshly matured seed (Fig. 2A). The enlargement of the seed, which resulted in the opening of both sides of the pericarp envelope, must therefore have been due to endosperm swelling, rather than to embryo growth. After the envelope had split open, the embryo began to grow (Fig. 2B) and developed from the heart to the torpedo stage (Fig. 2C). Following this, the embryo formed two well-developed cotyledons, and radicle protrusion occurred after a further 17 d (Fig. 2D), on average, by which time the E:S ratio had become  $0.48 \pm 0.07$ . After radicle emergence, the cotyledons continued to grow inside the seed, digesting the endosperm reserves (Fig. 2E, F), and emerged on average 23.5 d after the emergence of the radicle (Fig. 2G).

The minimum incubation period of > 100 d to radicle emergence clearly indicated the presence of seed dormancy in *A. trichopoda*. The absence of embryo development for at least the first 90 d of incubation indicated the presence of a physiological component to that dormancy, while the incompletely developed (heart-stage) embryo, following the breakage of PD, revealed the additional presence of MD. We therefore concluded that *A. trichopoda* must have a form of MPD. MPD is divided into the two principal sub-classes of ‘simple MPD’, in which the embryo requires warm ( $\geq 15$  °C) temperatures to grow and ‘complex MPD’ in which embryo requires cold (0–10 °C) temperatures to grow. Our preliminary observations clearly indicated the former scenario, and we therefore concluded the presence of a form of simple MPD in *A. trichopoda*.

#### Physiological components of dormancy

To elucidate the mechanistic basis of the PD component in *A. trichopoda* dormancy, we began by testing the water permeability of the mature drupe. When soaked in water for 72 h, the mass of both the DWFP and seed *sensu stricto* increased (Table 2), indicating that the pit was water permeable. Methylene blue solution entered the DWFP through the funiculus, opposite the hilum and embryo (Fig. 3), but stopped near

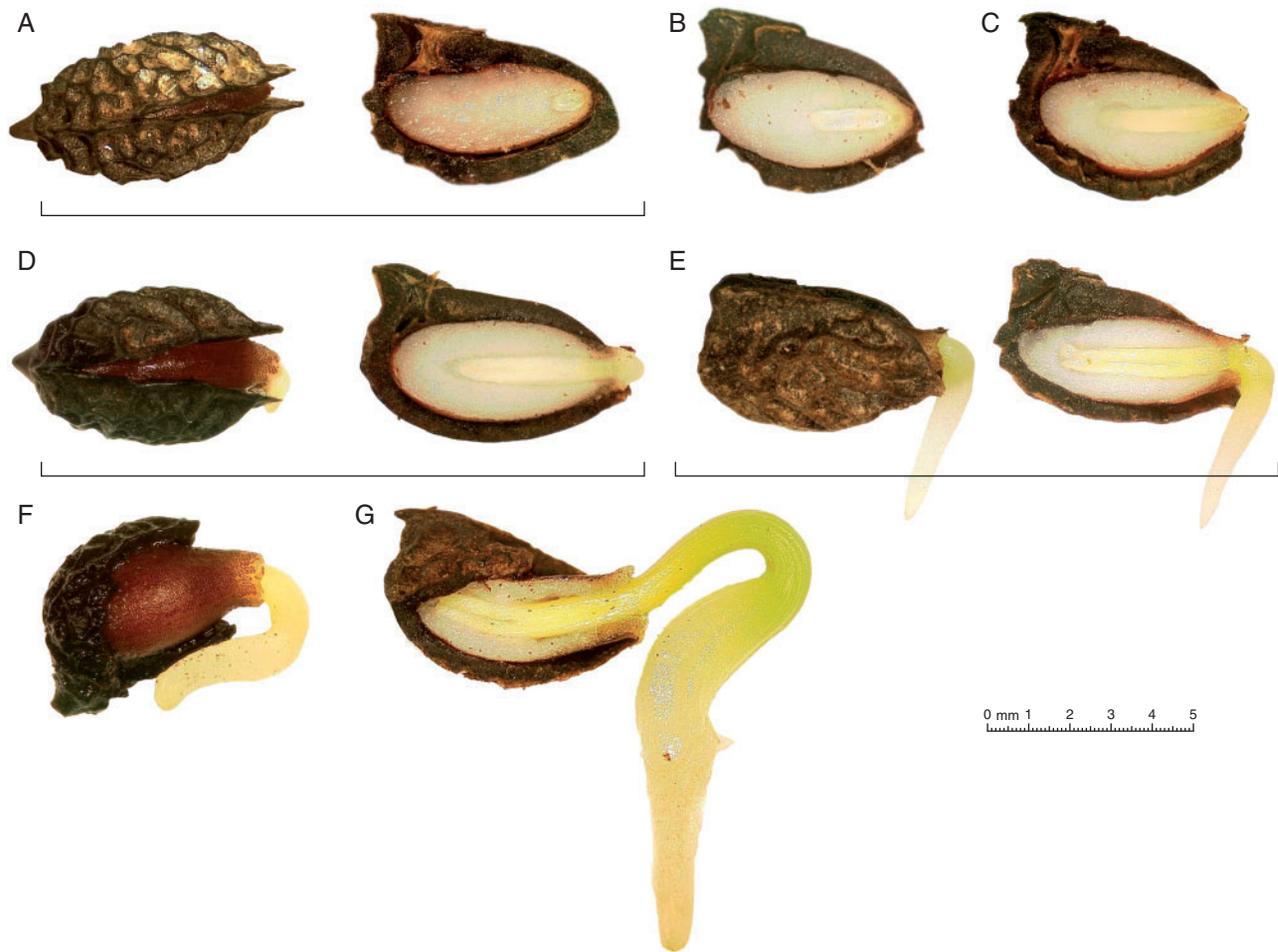


FIG. 2. (A) Reconstructed chronological sequence of *Amborella trichopoda* seed germination. Underlining indicates drupes without fleshy parts (DWFP) viewed whole or longitudinally cut. (A) Slightly open DWFP, whole (top view) and longitudinally cut (side view). (B and C) Embryos at later developmental stages, showing wider openings. (D) Whole and longitudinally cut DWFP at the stage of radicle emergence. (E–G) Subsequent developmental stages.

TABLE 2. Permeability of *Amborella trichopoda* drupes without fleshy part (DWFP) estimated by mass increase after 72 h, compared with controls, i.e. before imbibition

Part (mean of three replicates of 20 samples)	Mass of control (mg $\pm$ s.d.)	Mass after 72h (mg $\pm$ s.d.)	Mass increase (mg $\pm$ s.d.)	% increase
DWFP	428 $\pm$ 20	474 $\pm$ 20	47* $\pm$ 10	10.9 %*
Seed <i>sensu stricto</i>	114 $\pm$ 10	133 $\pm$ 10	18* $\pm$ 10	16.0 %*
Pit	313 $\pm$ 20	342 $\pm$ 20	28* $\pm$ 10	9.1 %*

Asterisks indicate statistically significant ( $P < 0.05$ ) increases in mass.

the hilum, possibly because the dye molecules were unable to enter the cells of the seed coat.

We therefore attempted to evaluate whether the PD component in *A. trichopoda* was due to mechanical resistance of the pericarp–mesocarp envelope, surrounding the seed. To do this, we compared the time course of germination in intact drupes, drupes scarified by acid treatment and seed *sensu stricto* that had been excised from all surrounding tissues (Fig. 4).

Scarification resulted in a reduction in the minimum germination time under greenhouse conditions from approx. 100 d to 60 d (Fig. 4). In agreement with this finding, a maximum of 70 % germination was reached after 138 d in scarified DWFP,

compared with 362 d in intact DWFP. Mechanical excision of the seed *sensu stricto* had an even greater effect on dormancy; excised seeds germinated in Petri dishes in a minimum of 18 d, compared with 62 d for non-scarified control DWFP. Maximum germination of 73 % was reached after 48 d for excised seed in Petri dishes, compared with 68 % after 342 d for non-scarified DWFP. Germination followed a logarithmic curve both for scarified DWFP in the greenhouse and excised seeds in Petri dishes. Taken together, these data clearly indicate that mechanical resistance of the pericarp–mesocarp envelope, external to the seed *sensu stricto*, makes a major contribution to the physiological component of simple MPD in *A. trichopoda*.

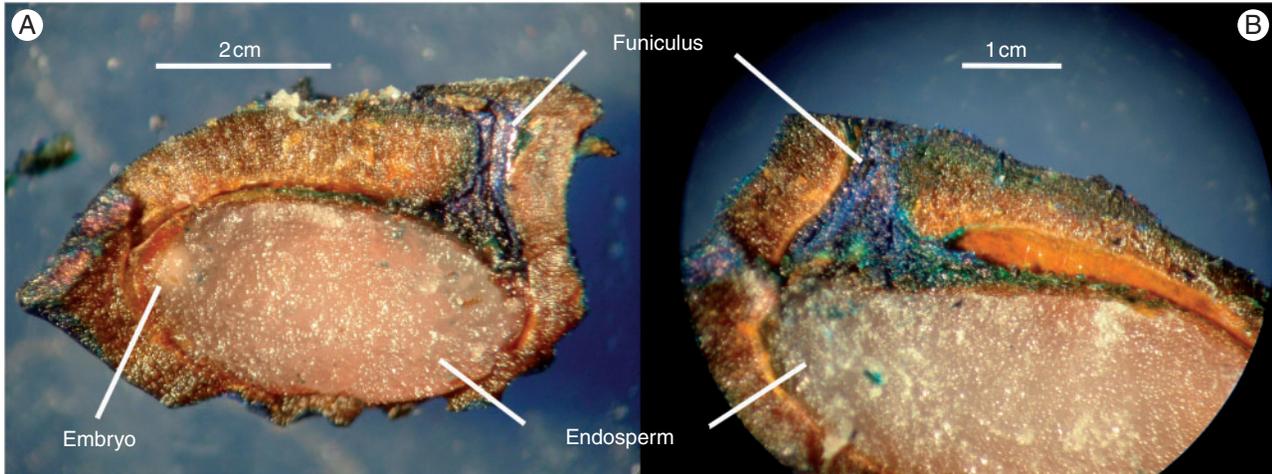


Fig. 3. Longitudinal section of a drupe without its fleshy part (DWFP) soaked for 72h in 1 % methylene blue solution. (A) Entire DWFP. (B) Zoom near the funiculus.

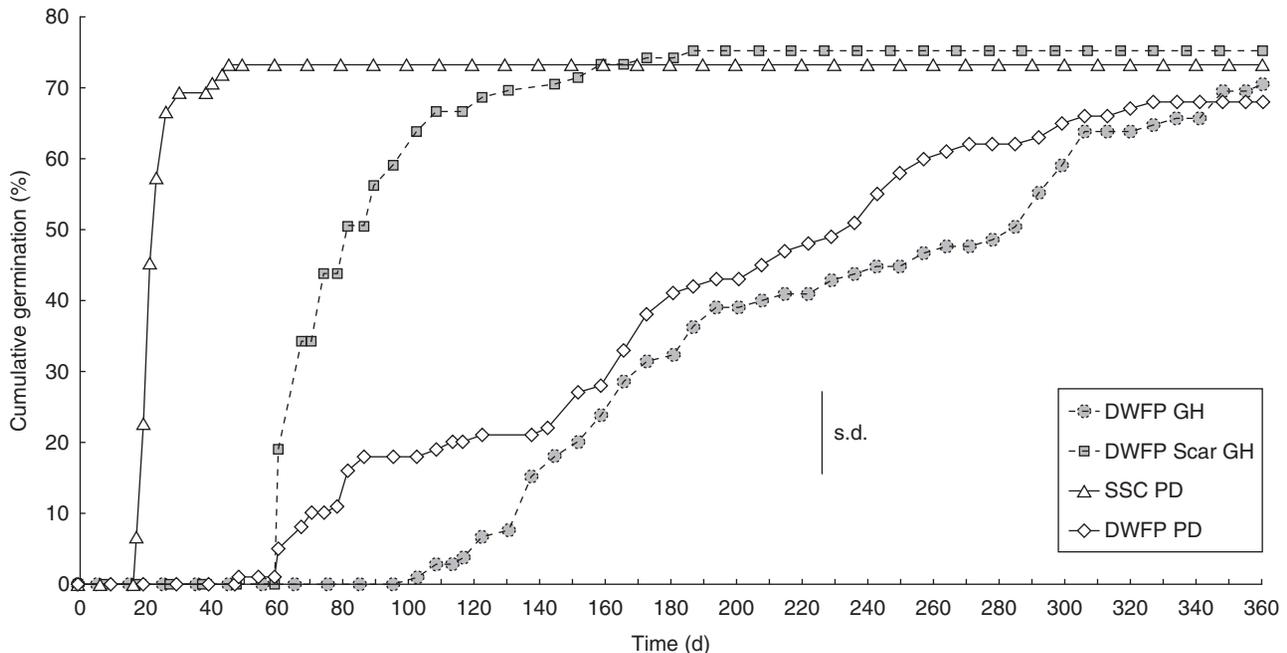


Fig. 4. Time course of germination for *Amborella trichopoda* seeds. Seeds *sensu stricto* (SSC PD) and drupes without fleshy part (DWFP PD) in Petri dishes at  $25 \pm 1^\circ\text{C}$  with continuous light, and for DWFP with (DWFP GH) or without (DWFP Scar GH) acid scarification on trays at  $25 \pm 4^\circ\text{C}$  in a greenhouse at high humidity. Means were derived from four replicates of 25 seeds *sensu stricto* or DWFP. The vertical bar denotes the largest s.d.

The period of time (approx. 90 d) required to break the physiological component of MPD in *A. trichopoda* is closely comparable with that in numerous species displaying non-deep simple MPD, rather than the alternative deep simple MPD (Baskin and Baskin, 2014), suggesting the presence of the former dormancy type. To confirm this hypothesis, we used a discriminatory test using exogenous  $\text{GA}_3$ . The percentages of germination after 143 d and the rapidity of germination of non-scarified DWFP were significantly increased by incubation in  $\text{GA}_3$  solutions of 50 or 100 ppm, though higher concentrations of 500 or 1000 ppm  $\text{GA}_3$  were inhibitory to germination

(Fig. 5). These data, taken together with our observations that seeds only require incubation at temperatures of  $\geq 15^\circ\text{C}$  for dormancy break and germination, confirmed the presence of non-deep PD in seeds of *A. trichopoda* and thus the presence of non-deep simple MPD.

#### Test for epicotyl MPD

Six distinct sub-classes of simple MPD have been recognized (Baskin and Baskin, 2004), four of which include specific forms of epicotyl dormancy. To evaluate the presence of epicotyl

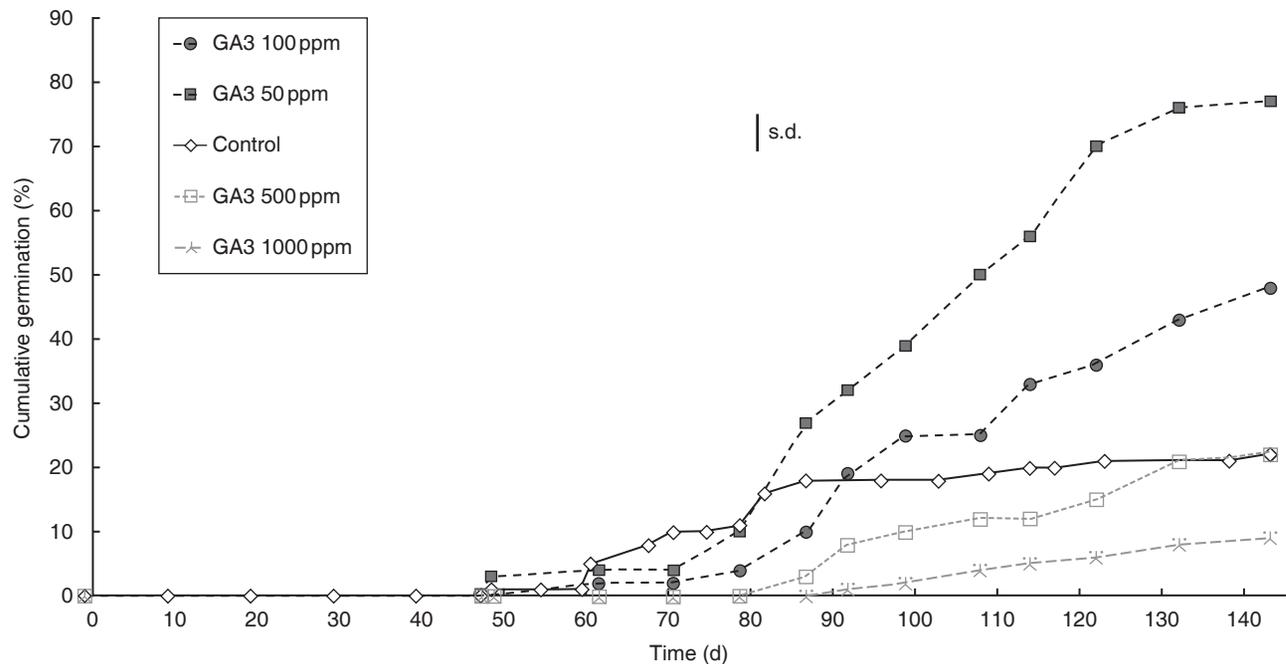


FIG. 5. Time course of germination for non-scarified *Amborella trichopoda* drupes without fleshy parts (DWFP) in Petri dishes. DWFP were incubated at  $25 \pm 1^\circ\text{C}$  with continuous light on germination paper imbibed with solutions of 0, 50, 100, 500 and 1000 ppm of  $\text{GA}_3$ . Means were derived from four replicates of 25 DWFP. The vertical bar denotes the largest s.d.

TABLE 3. Time required for radicle and cotyledon emergence from *Amborella trichopoda* seeds and the effects on this process of  $\text{GA}_3$

Treatment (24 h incubation)	Mean no. of days between pit opening and radicle emergence	Sub-treatment on 5 mm long rootlets	Mean no. of days between radicle protrusion and cotyledon emergence
Water ( $n = 98$ )	17.6* ( $n = 95$ )	Untreated control ( $n = 41$ ) $\text{GA}_3$ in lanolin ( $n = 38$ )	23.5* ( $n = 39$ ) 24.6* ( $n = 37$ )
50 ppm $\text{GA}_3$ ( $n = 100$ )	15.5* ( $n = 98$ )	–	–

Asterisks indicate statistically significant ( $P < 0.05$ ) differences between the treatments shown in each column (50 ppm  $\text{GA}_3$  vs. water and  $\text{GA}_3$  in lanolin vs. untreated controls, respectively).

Numbers of replicates ( $n$ ) in each treatment group are indicated in parentheses.

$\text{GA}_3$  in lanolin treatments and their respective controls were performed on 79 surviving seedlings ( $n = 41$  and 38, respectively) from the group of 98 seeds previously incubated in water only.

dormancy in *A. trichopoda*, we measured the time necessary for cotyledon emergence, following radicle emergence. Cotyledon emergence from non-scarified DWFP occurred rapidly after radicle emergence, from 15 d for the earliest appearance of cotyledons to 30 d for the latest, giving an overall mean of 23.5 d (Table 3). The application of 50 ppm  $\text{GA}_3$  in lanolin paste had no significant effect on the speed of this process ( $P < 0.05$ ). There are two sub-classes of simple MPD which do not involve epicotyl dormancy: deep simple MPD and non-deep simple MPD. As our findings indicate the absence of epicotyl dormancy in *A. trichopoda*, these are consistent with the identification in this species (above) of the non-deep simple level of MPD.

#### Character reconstruction by parsimony

Character-state mapping indicates that the MRCA of living angiosperms possessed MPD and reveals a transition to PD

within Nymphaeales. We combined the identification of the level of MPD in *A. trichopoda* with published data on the class of seed dormancy in the three most basal lineages of flowering plants, Amborellales, Nymphaeales and Austrobaileyales. Notably, this analysis included data on three species of *Trithuria* from the basally diverging family Hydatellaceae (Tuckett *et al.*, 2010), not included in previous reconstructions on the evolution of dormancy (Willis *et al.*, 2014). We also included in our analysis the likely dormancy type in the MRCA of the remaining angiosperm lineage, the euangiosperms, which was inferred by Willis *et al.* (2014) as ambiguous between MPD and PD.

We investigated the evolution of dormancy mechanisms in the angiosperms using the consensus view of angiosperm phylogeny (Byng *et al.*, 2016) in which Amborellales alone is sister to all other living angiosperms, (Fig. 6A, B), and the alternative view (Qiu *et al.*, 2001, 2010; Xi *et al.*, 2014) in which a clade

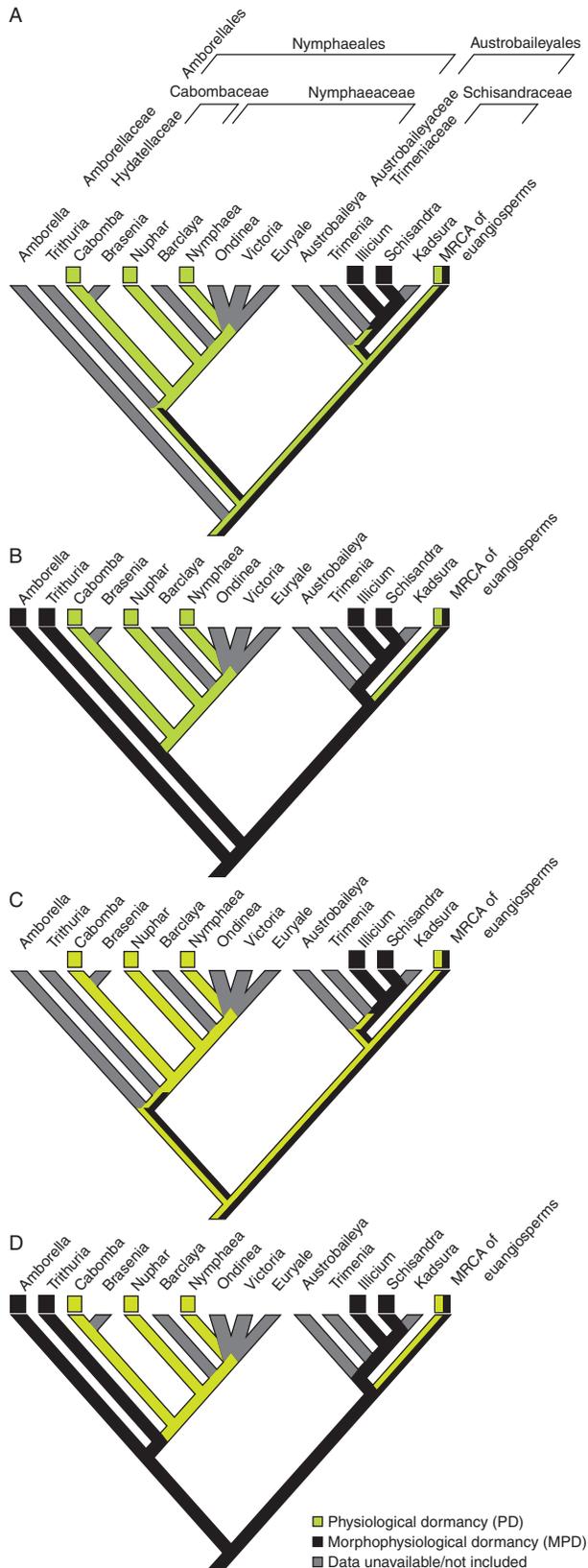


Fig. 6. Mapping of germination classes by parsimony onto two alternative topologies for the base of angiosperm phylogeny. Alternative phylogenies are considered in which either Amborellales alone (A and B), or Amborellales +

consisting of Amborellales + Nymphaeales is sister to all other living angiosperms (Fig. 6C, D). These analyses included the dormancy type in the MRCA of euangiosperms, as reconstructed by Willis *et al.* (2014). Therefore, all living ANA-grade genera, and a very wide sampling of euangiosperms (previously performed by Willis *et al.*, 2014), are represented in these analyses. Parsimony reconstruction, omitting data from *A. trichopoda* and *Trithuria* spp., failed to resolve the dormancy type at the base of the flowering plants in both of the angiosperm topologies considered (Fig. 6A, C). However, the addition of data from these taxa unambiguously resolved the dormancy type at the base of the angiosperm tree in both topologies considered (Fig. 6B, D). Using either of these alternative evolutionary scenarios, the base of the living angiosperms was resolved unambiguously as possessing MPD. We therefore conclude MPD as the likely plesiomorphic condition of the living angiosperms. From our reconstruction, it appears that the PD present in Nymphaeaceae and Cabombaceae arose through the loss of the morphological component of dormancy from MPD that was present in the MRCA of all living Nymphaeales (Fig. 6B, D).

## DISCUSSION

### *Multiple significance of seed dormancy studies in Amborella trichopoda*

In this study, we have determined the precise type of seed dormancy present in *A. trichopoda*, the only known representative of Amborellales and likely sister to all other flowering plants (Figs 1–5). We conclude that seeds of this species have non-deep simple MPD, in which the PD component is broken before the MD component (and is, therefore, ‘simple’; Fig. 2), and in which embryo growth occurred at high temperatures and GA<sub>3</sub> was effective in promoting germination (Fig. 4). The PD component in *A. trichopoda* depends heavily on mechanical resistance to embryo growth from the pericarp–mesocarp envelope of the drupe (Fig. 4). Interestingly, we reveal the presence of an amorphous silicate-crystal layer, situated between the endocarp and mesocarp (Fig. 1E), which may make a significant contribution to the mechanical properties of the drupe, and therefore to the physiological component of dormancy in *A. trichopoda*.

The significance of our results is 3-fold. First, no character-state mapping at the base of the living angiosperms can be complete without the inclusion of data from *A. trichopoda*. Our study has thus enabled, for the first time, the precise reconstruction of the evolution of dormancy classes in the MRCA of living angiosperms, which we conclude to have possessed MPD (Fig. 6). This conclusion is useful for the interpretation of the

Nymphaeales (C and D) are sister to all other living angiosperms. When dormancy data from *Amborella* and *Trithuria* are excluded (A and C), the analysis fails to resolve the dormancy class at the base of living angiosperms (red arrows) between PD and MPD. The inclusion of dormancy data from *Amborella* and *Trithuria* (B and D) results in the resolution of the basal character state in the living angiosperms to MPD for both phylogenetic topologies considered. A transition from MPD to PD (blue arrows) can be inferred within Nymphaeales, in a common ancestor of Cabombaceae and Nymphaeaceae, in either phylogenetic topology (B and D). Classification of genera into families and orders is shown in (A).

subsequent evolution of dormancy classes within the angiosperms, such as the transition from MPD to PD that we infer to have occurred in a common ancestor of Nymphaeaceae and Cabombaceae (Fig. 6). Secondly, our findings contribute to a growing picture of the ecophysiology of *A. trichopoda* within its endemic habitat of the New Caledonian rain forest. We thus add data on seed dormancy and germination to studies of photosynthesis (Feild *et al.*, 2001), stomatal function (Rudall and Knowles, 2013), vascular function (Feild *et al.*, 2000; Turgeon and Medville, 2011; Feild and Wilson, 2012), flowering time (Fourcade *et al.*, 2015), pollination biology (Thien *et al.*, 2009), seed physiology (Poncet *et al.*, 2015), population structure (Thien *et al.*, 2003; Poncet *et al.*, 2013) and genetic diversity (Poncet *et al.*, 2012). These diverse studies of *A. trichopoda* make a major contribution to the concerted reconstruction of the ecophysiological features of the first flowering plants (see Feild *et al.*, 2004; Feild and Arens, 2007). Thirdly, the investigation of dormancy in *A. trichopoda* is of practical use both for species conservation (e.g. for use in seed banks) and for the exploitation of *A. trichopoda* as an experimental model representing ANA-grade angiosperms. Such experimental uses may require the manipulation of dormancy and germination to facilitate numerous procedures including genetic crosses, tissue culture and plant transformation (see Scutt and Vandebussche, 2014).

*The conclusion that morphophysiological dormancy was present at the base of the living angiosperms is robust and should withstand any reasonably foreseeable change to angiosperm or gymnosperm phylogeny*

The recently published study by Willis *et al.* (2014) concluded the presence of MPD in the MRCA of all living seed plants (angiosperms + gymnosperms) and indicated that MPD was the probable dormancy class near the base of living angiosperms. However, the principal conclusions of Willis *et al.* (2014) may have depended heavily on the version of gymnosperm phylogeny included in their analysis. The phylogenetic topology used by Willis *et al.* (2014) included Cycadales, followed by Gingoales, as the two most basal clades of living gymnosperms, and placed Gnetales in sister position to all conifers. This placement of Gnetales is consistent with several published phylogenies (e.g. Chaw *et al.*, 1997; Wickett *et al.*, 2014), though others place Gnetales within conifers, as sister to Pinaceae (e.g. Bowe *et al.*, 2000; Zhong *et al.*, 2010), or as sister to remaining conifers, excluding Pinaceae (e.g. Nickrent *et al.*, 2000; Doyle, 2006). Importantly, two gymnosperm phylogenies have recently been proposed in which Gnetales are placed as sister to all other gymnosperms (de la Torre-Barcelona *et al.*, 2009; Lee *et al.*, 2011). As all three monotypic families within Gnetales possess PD, rather than MPD (summarized by Willis *et al.*, 2014), the phylogenetic placement of Gnetales is likely to have a critical impact on the reconstruction of dormancy mechanisms at the base of gymnosperms and, by extension, at the base of the living seed plants.

In contrast to the situation in gymnosperms, the basal nodes of angiosperm phylogeny, as explained above, have been broadly established, with only minor differences between a consensus view supported by the majority of studies (Byng *et al.*,

2016), and an alternative view supported by a smaller number of studies (Qiu *et al.*, 2001, 2010; Xi *et al.*, 2014). We therefore preferred, in the present work, to reconstruct the type of dormancy of the MRCA of all living angiosperms using data exclusively from angiosperms. We performed four analyses (Fig. 6), including both of the alternative evolutionary scenarios that emerge from the current literature, in each case with and without data from *A. trichopoda* (from this work) and *Trithuria* spp. (Tuckett *et al.*, 2010), which were not available to, or not included in, previous evolutionary reconstructions. We show that the addition of data from these key basal taxa leads to the resolution of the MPD in the MCRA of living angiosperms, regardless of which of the two phylogenetic topologies that we analysed is considered. We do not, therefore, expect this evolutionary conclusion to be brought into question by any reasonably foreseeable change to the consensus view of angiosperm phylogeny. In addition, as our principal evolutionary conclusion was reached using well-established data exclusively from angiosperms, it should not be affected by the current flux and uncertainty in gymnosperm phylogeny.

Importantly, our conclusion that MPD was the likely dormancy class in the first angiosperms supports the model proposed by Willis *et al.* (2014) for the evolution within angiosperms of PD and MD, via alternative routes, from MPD. The establishment of MPD at the base of the angiosperms also supports a second hypothesis advanced by Willis *et al.* (2014) that key ancestral species within the euangiosperms, in which PD had already evolved from MPD, then acted as foci for the evolutionary diversification of seed dormancy mechanisms.

#### ACKNOWLEDGEMENTS

We acknowledge grant funding from to the Fondation Ars-Cuttoli-Paul Appell under the aegis of the Fondation de France. We are also grateful to the Province Sud de Nouvelle-Calédonie for permission for seed collection and for additional financial support. We thank Jacques Rabier, Roger Notonier and Alain Tonetto at the SCME, University of Provence, France for help with SEM, and finally special thanks to C. Zongo (deceased) for field collection of seeds.

#### LITERATURE CITED

- Bailey IW, Swamy BGL. 1948. *Amborella trichopoda* Baill., a new morphological type of vesselless dicotyledon. *Journal of the Arnold Arboretum* **29**: 245–254. + 5 plates.
- Baskin CC, Baskin JM. 2014. *Seeds: ecology, biogeography, and evolution of dormancy and germination*, 2nd edn. San Diego: Elsevier/Academic Press.
- Baskin JM, Baskin CC. 2004. A classification system for seed dormancy. *Seed Science Research* **14**: 1–16.
- Bobrov AVFC, Endress PK, Melikian AP, Romanov MS, Sorokin AN, Bejerano AP. 2005. Fruit structure of *Amborella trichopoda* (Amborellaceae). *Botanical Journal of the Linnean Society* **148**: 265–274.
- Bowe LM, Coat G, de Pamphilis CW. 2000. Phylogeny of seed plants based on all three genomic compartments: extant gymnosperms are monophyletic and Gnetales' closest relatives are conifers. *Proceedings of the National Academy of Sciences, USA* **97**: 4092–4097.
- Bewley JD, Bradford K, Hilhorst H, Nonogaki H. 2013. *Seeds: physiology of development, germination and dormancy*, 3rd edn. Springer editions.
- Byng JW, Chase MW, Christenhusz MJM *et al.* 2016. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. *Botanical Journal of the Linnean Society* **181**: 1–20.

- Chaw SM, Zharkikh A, Sung HM, Lau TC, Li WH. 1997. Molecular phylogeny of extant gymnosperms and seed plant evolution: analysis of nuclear 18S rRNA sequences. *Molecular Biology and Evolution* **14**: 56–68.
- Doyle JA. 2006. Seed ferns and the origin of angiosperms. *Journal of the Torrey Botanical Society* **133**: 169–209.
- Feild TS, Arens NC. 2007. The ecophysiology of early angiosperms. *Plant, Cell & Environment* **30**: 291–309.
- Feild TS, Wilson JP. 2012. Evolutionary voyage of angiosperm vessel structure–function and its significance for early angiosperm success. *International Journal of Plant Sciences* **173**: 596–609.
- Feild TS, Zweiniecki MA, Brodribb T, Jaffre T, Donoghue MJ, Holbrook NM. 2000. Structure and function of tracheary elements in *Amborella trichopoda*. *International Journal of Plant Sciences* **161**: 705–712.
- Feild TS, Brodribb T, Jaffre T, Holbrook NM. 2001. Acclimation of leaf anatomy, photosynthetic light use, and xylem hydraulics to light in *Amborella trichopoda* (Amborellaceae). *International Journal of Plant Sciences* **162**: 999–1008.
- Feild TS, Arens NC, Doyle JA, Dawson TE, Donoghue MJ. 2004. Dark and disturbed: a new image of early angiosperm ecology. *Paleobiology* **30**: 82–107.
- Friedman WE, Bachelier JB, Hormaza JI. 2012. Embryology in *Trithuria submersa* (Hydatellaceae) and relationships between embryo, endosperm, and perisperm in early-diverging flowering plants. *American Journal of Botany* **99**: 1083–1085.
- Fourcade F, Pouteau R, Jaffre T, Marmey P. 2015. *In situ* observations of the basal angiosperm *Amborella trichopoda* reveal a long fruiting cycle overlapping two annual flowering periods. *Journal of Plant Research* **128**: 821–828.
- ISTA. 1999. International rules for seed testing. *Seed Science and Technology* 27 (supplement).
- Lee EK, Cibrian-Jaramillo A, Kolokotronis S-O, et al. 2011. A functional phylogenomic view of the seed plants. *PLoS Genetics* **7**: e1002411.
- Linkies A, Graeber K, Knight C, Leubner-Metzger G. 2010. The evolution of seeds. *New Phytologist* **186**: 817–831.
- Löhne C, Borsch T, Wiersema JH. 2007. Phylogenetic analysis of Nymphaeales using fast-evolving and noncoding chloroplast markers. *Botanical Journal of the Linnean Society* **154**: 141–163.
- MacKay AC, McGill CR, Fountain DW, Southward RC. 2002. Seed dormancy and germination of a panel of New Zealand plants suitable for revegetation. *New Zealand Journal of Botany* **40**: 373–382.
- Maddison WP, Maddison DR. 2015. *Mesquite: a modular system for evolutionary analysis. Version 3.04*. <http://mesquiteproject.org>.
- Nickrent DL, Parkinson CL, Palmer JD, Duff RJ. 2000. Multigene phylogeny of land plants with special reference to bryophytes and the earliest land plants. *Molecular Biology and Evolution* **17**: 1885–1895.
- Orozco-Segovia A, Marquez-Guzman J, Sanchez-Coronado ME, De Buen AG, Baskin JM, Baskin CC. 2007. Seed anatomy and water uptake in relation to seed dormancy in *Opuntia tomentosa* (Cactaceae, Opuntioideae). *Annals of Botany* **99**: 581–592.
- Poncet V, Couderc M, Tranchant-Dubreuil C, et al. 2012. Microsatellite markers for *Amborella* (Amborellaceae), a monotypic genus endemic to New Caledonia. *American Journal of Botany* **99**: E411–E414.
- Poncet V, Munoz F, Munzinger J, et al. 2013. Phylogeography and niche modelling of the relict plant *Amborella trichopoda* (Amborellaceae) reveal multiple Pleistocene refugia in New Caledonia. *Molecular Ecology* **22**: 6163–6178.
- Poncet V, Scutt C, Tournebize R, et al. 2015. The *Amborella* vacuolar processing enzyme family. *Frontiers in Plant Science* **6**: 618.
- Qiu YL, Lee J, Whitlock BA, Bernasconi-Quadroni F, Dombrowska O. 2001. Was the ANITA rooting of the angiosperm phylogeny affected by long-branch attraction? *Molecular Biology and Evolution* **18**: 1745–1753.
- Qiu Y-L, Li L, Wang B, et al. 2010. Angiosperm phylogeny inferred from sequences of four mitochondrial genes. *Journal of Systematics and Evolution* **48**: 391–425.
- Rabier J, Laffont-Schwob I, Notonier R, Fogliani B, Bouraïma-Madjèbi S. 2008. Anatomical element localization by EDXS in *Grevillea exul* var. *exul* under nickel stress. *Environmental Pollution* **156**: 1156–1163.
- Rudall PJ, Knowles EVW. 2013. Ultrastructure of stomatal development in early-divergent angiosperms reveals contrasting patterning and pre-patterning. *Annals of Botany* **112**: 1031–1043.
- Saarela JM, Rai HS, Doyle JA, et al. 2007. Hydatellaceae identified as a new branch near the base of the angiosperm phylogenetic tree. *Nature* **446**: 312–315.
- Scutt CP, Vandenbussche M. 2014. Current trends and future directions in flower development research. *Annals of Botany* **114**: 1399–1406.
- Thien LB, Sage TL, Jaffre T, et al. 2003. The population structure and floral biology of *Amborella trichopoda* (Amborellaceae). *Annals of the Missouri Botanical Garden* **90**: 466–490.
- Thien LB, Bernhardt P, Devall MS, et al. 2009. Pollination biology of basal angiosperms (ANITA grade). *American Journal of Botany* **96**: 166–182.
- Tobe H, Jaffré T, Raven PH. 2000. Embryology of *Amborella* (Amborellaceae): descriptions and polarity of character states. *Journal of Plant Research* **113**: 271–280.
- de la Torre-Barcelona JE, Kolokotronis S-O, Lee EK, et al. 2009. The impact of outgroup choice and missing data on major seed plant phylogenetics using genome-wide EST data. *PLoS One* **4**: e5764.
- Tuckett RE, Merritt DJ, Rudall PJ, et al. 2010. A new type of specialized morphophysiological dormancy and seed storage behaviour in Hydatellaceae, an early-divergent angiosperm family. *Annals of Botany* **105**: 1053–1061.
- Turgeon R, Medville R. 2011. *Amborella trichopoda*, plasmodesmata, and the evolution of phloem loading. *Protoplasma* **248**: 173–180.
- Wickett NJ, Mirarab S, Nguyen N, et al. 2014. Phylotranscriptomic analysis of the origin and early diversification of land plants. *Proceedings of the National Academy of Sciences, USA* **111**: E4859–E4868.
- Willis CG, Baskin CC, Baskin JM, et al. 2014. The evolution of seed dormancy: environmental cues, evolutionary hubs, and diversification of the seed plants. *New Phytologist* **203**: 300–309.
- Xi Z, Liu L, Rest JS, Davis CC. 2014. Coalescent versus concatenation methods and the placement of *Amborella* as sister to water lilies. *Systematic Biology* **63**: 919–932.
- Zhong B, Yonezawa T, Zhong Y, Hasegawa M. 2010. The position of Gnetales among seed plants: overcoming pitfalls of chloroplast phylogenomics. *Molecular Biology and Evolution* **27**: 2855–2863.